

doi: 10.13241/j.cnki.pmb.2015.10.006

糖尿病大鼠 ghrelin 和 nesfatin-1 表达动力学及其调控 *

王丽霞^{1,2} 王巧玲¹ 逢明杰³ 祝海³ 郭菲菲¹ 孙向荣¹ 徐珞^{1△}

(1 青岛大学医学院病理生理学教研室 山东青岛 266021;

2 鄄城县人民医院 山东菏泽 274700;3 青岛市立医院 山东青岛 266000)

摘要 目的:探讨 STZ 诱导糖尿病大鼠 ghrelin 和 nesfatin-1 动力学及分泌调节变化。方法:STZ 诱导糖尿病大鼠模型;采用葡萄糖脱氢酶分析法测量血浆葡萄糖水平;免疫放射分析检测血浆 ghrelin、nesfatin-1、胰岛素、胰岛素样生长因子 1(IGF-1)、生长激素(GH)含量;采用 real-time PCR 检测 ghrelin mRNA 水平变化;免疫组化观察 ghrelin 和 nesfatin-1 免疫活性细胞数量。结果:糖尿病大鼠体重显著降低($t=23.16, P<0.01$),血糖水平显著升高($t=22.55, P<0.01$),血浆胰岛素和 IGF-1 水平显著降低($t=6.50, t=24.13, P<0.01$),但 GH 水平显著升高($t=3.30, P<0.05$)。糖尿病大鼠血浆总 ghrelin($t=7.03, P<0.01$)和活性 ghrelin($t=3.33, P<0.05$)水平均显著升高,血浆 nesfatin-1 水平则显著降低($t=6.24, P<0.01$);糖尿病大鼠血浆总 ghrelin 与 GH($r=0.81, P<0.01$)和 IGF-1 水平($r=-0.58, P<0.01$)呈显著相关性;与对照组大鼠相比,糖尿病大鼠胃总 ghrelin($t=16.86, P<0.01$)和活性 ghrelin($t=3.30, P<0.05$)水平均显著降低;而胃 nesfatin-1($t=7.93, P<0.01$)水平则显著升高。胃总 ghrelin 水平与血浆 IGF-1 水平呈明显相关性($r=0.65, P<0.01$);与对照组大鼠相比,糖尿病大鼠胃 ghrelin mRNA 表达水平显著升高($t=16.8, P<0.01$),胃底 ghrelin 免疫活性细胞数量显著减少($t=3.98, P<0.01$);实验中给予大鼠自由饮食,糖尿病大鼠血浆总 ghrelin 水平显著增加($t=7.53, P<0.01$),nesfatin-1 水平显著降低($t=5.46, P<0.01$)。糖尿病大鼠注射胰岛素后,可使增加的 ghrelin 水平($t=1.76, P=0.11$)和降低的 nesfatin-1 水平接近正常($t=1.96, P=0.06$);且胰岛素可显著反转糖尿病大鼠胃总 ghrelin($t=8.54, P<0.01$)和 nesfatin-1 水平($t=2.42, P<0.05$);以及注射胰岛素后,糖尿病大鼠胃底 ghrelin 细胞显著增加,nesfatin-1 细胞明显减少($t=3.21, t=2.59, P<0.05$)。结论:Ghrelin 或 nesfatin-1 参与糖尿病大鼠能量平衡调控。

关键词:Ghrelin; Nesfatin-1; 糖尿病; 胃; 生长激素

中图分类号:Q95-3;R587.1 文献标识码:A 文章编号:1673-6273(2015)10-1820-05

Dynamics Expression and Regulation of Ghrelin and Nesfatin-1 in Diabetic Rats*

WANG Li-xia^{1,2}, WANG Qiao-ling¹, PANG Ming-jie³, ZHU Hai³, GUO Fei-fei¹, SUN Xiang-rong¹, XU Luo^{1△}

(1 Dept. of Pathophysiology, Medical College of Qingdao University, Qingdao, Shandong, 266021, China; 2 Yun Cheng people's hospital, Heze, Shandong, 274700, China; 3 Qingdao Municipal Hospital, Qingdao, Shandong, 266000, China)

ABSTRACT Objective: To study the changes of ghrelin and nesfatin-1 dynamics and secretory regulation in rats with STZ-induced diabetes. **Methods:** Diabetes was induced by intraperitoneal injection of streptozotocin (STZ). Serum glucose were measured by the glucose dehydrogenase, while the levels of ghrelin, nesfatin-1, insulin, IGF-1 and GH were detected by RIA. Preproghrelin mRNA level was tested using real-time RT-PCR and the ghrelin and nesfatin-1-immunoreactive cells were observed using immunohistochemistry. **Results:** The weight of diabetic rats decreased significantly ($t=23.16, P<0.01$), the glucose was increased significantly ($t=22.55, P<0.01$), plasma insulin and IGF-1 level was decreased significantly ($t=6.50, t=24.13, P<0.01$), but the GH level was increased significantly ($t=3.30, P<0.05$). The level of total ($t=7.03, P<0.01$) and active ($t=3.33, P<0.05$) plasma ghrelin of the diabetic rats was increased significantly, while the plasma nesfatin-1 level was decreased ($t=6.24, P<0.01$), compared with that of the control group. The total plasma ghrelin level of the diabetic rats correlated significantly with their serum GH level ($r=0.81, P<0.01$) and with their serum IGF-1 level ($r=-0.58, P<0.01$). The gastric total ($t=16.86, P<0.01$) and active ($t=3.30, P<0.05$) ghrelin level of the diabetic rats was decreased significantly, while the gastric nesfatin-1 levels was increased ($t=7.93, P<0.01$) compared with that in the control group, and their gastric total ghrelin level correlated significantly with their serum IGF-1 level ($r=0.65, P<0.01$). The preproghrelin mRNA expression in the stomach was increased significantly ($t=16.8, P<0.01$) in the diabetic rats compared with the control and the number of ghrelin-immunoreactive cells in the gastric fundus of the diabetic rats was decreased significantly ($t=3.98, P<0.01$). Giving rats free diet in the experiments, the total plasma ghrelin level in diabetic rats was increased significantly ($t=7.53, P<0.01$), nesfatin-1 level

* 基金项目:国家自然科学基金项目(31071014,81100260,81270460,81300281,81470815);

青岛市科技局项目(13-1-4-170-jch,11-2-3-3-(2)-nsh,14-2-3-3-nsh)

作者简介:王丽霞(1979-),女,硕士研究生,主治医师,主要研究方向:神经内分泌,电话:0532-82991713,E-mail: sszx179@sina.com

△通讯作者:徐珞,E-mail: xu.luo@163.com

(收稿日期:2014-10-29 接受日期:2014-11-25)

significantly reduced ($t=5.46$, $P<0.01$)。After insulin injections, the increasing level of ghrelin ($t=1.76$, $P=0.11$) and reducing level of nesfatin-1 in diabetic rats was close to normal ($t=1.96$, $P=0.06$)。Insulin significantly reversed the total gastric ghrelin ($t=8.54$, $P<0.01$) and nesfatin-1 level ($t=2.42$, $P<0.05$) of diabetes rats。After injection of insulin, ghrelin cells were increased significantly, while nesfatin-1 cells were decreased significantly ($t=3.21$, $t=2.59$, $P<0.05$) in stomach fundus of diabetic rats。Conclusions: Ghrelin or nesfatin-1 may participate in the regulation of energy balance in diabetic rats。

Key words: Ghrelin; Nesfatin-1; Diabetes; Stomach; Growth hormone

Chinese Library Classification (CLC): Q95-3; R587.1 Document code: A

Article ID: 1673-6273(2015)10-1820-05

前言

Ghrelin 是从人类和大鼠胃中分离的一种新型生长激素促分泌素^[1], 在其第 3 位丝氨酸残基上有特异性 N 端辛酰基修饰, 对 ghrelin 的生理特性至关重要。Ghrelin 的主要生理功能是促进人类及啮齿类动物生长激素分泌, 促进摄食^[2]及胃运动^[3]。负能量平衡状态比如饥饿或恶病质、神经性厌食症所致的低体重可使血浆 ghrelin 水平增加^[4-8]。许多组织可表达前体 ghrelin mRNA^[9], 其中胃底表达水平最高^[10]。Nesfatin-1 是 2006 年发现的一种新型厌食肽^[11], 在中枢、外周胃肠、胰岛等部位均有表达, 具有减少摄食、抑制胃肠运动等作用。据报道胃底 A 样细胞可分泌 ghrelin 和 nesfatin-1。

链脲佐菌素(STZ)是一种抗生素, 可通过破坏胰腺胰岛内分泌细胞诱导糖尿病生成。STZ 诱导糖尿病大鼠可出现体重降低、多食、血糖升高、低胰岛素血症^[12,13]。至今还没有报道 STZ 诱导胰岛素依赖型糖尿病大鼠 ghrelin 和 nesfatin-1 的动力学变化。本文应用免疫放射分析法 (RIA)、实时酶联反应法 (RT-PCR) 及免疫组织化学技术等方法, 探讨 STZ 诱导糖尿病大鼠 ghrelin 和 nesfatin-1 动力学及分泌调节变化。

1 材料和方法

1.1 实验动物

8 周龄雄性 Wistar 大鼠 45 只, 随机分为 3 组: 对照组 ($n=15$), 腹腔注射生理盐水; STZ 诱导糖尿病大鼠组 ($n=15$), 腹腔注射 STZ (60 mg/kg) 诱导糖尿病; 胰岛素处理糖尿病大鼠组 ($n=15$), 在注射 STZ 一周后给予皮下注射中性鱼精蛋白人胰岛素 (7 U/只)。所有动物自由饮水和进食, 4 周后大鼠禁食 16 h, 乙醚麻醉处死。分离大鼠胃、十二指肠和上端结肠, 生理盐水冲洗。

1.2 血液及相关激素分析

采用葡萄糖脱氢酶分析法测量血浆葡萄糖水平, 放射免疫分析 (RIA) 参照文献^[14,15] 进行, 检测血浆胰岛素、胰岛素样生长因子 1 (IGF-1)、生长激素 (GH) 以及血浆和胃组织 ghrelin、nesfatin-1 水平。

1.3 Real-time PCR

使用 Tri-RNA 试剂盒提取下丘脑组织总 RNA, 用 1 μ g 总 RNA 和寡核苷酸 dT 引物合成 cDNA。cDNA 与 SYBR Premix Ex TaqII 和特定引物混合, 进行 RT-PCR: 95° C 3 s, 60° C 30 s, 50 个循环。Ghrelin 引物: 5' -GGA ATC CAA GAA GCC ACC AGC-3'; 5' -GCT CCT GACAGC TTG ATG CCA-3'。Ghrelin 探针序列: 5' -FAM-AAC TGC AGC CAC GAG CTC TGG AAG GC-TAMRA-3'。内源性对照 GAPDH 引物: 5'

-TTC AAC GGC ACA GTC AAG GC-3'; 5' -GCC TTC TCC ATG GTG GTG AAG-3'。GAPDH 探针序列: 5' -FAM-CCC ATC ACC ATC TTC CAG GAG CGA GA-TAMRA-3'。

1.4 免疫组织化学染色

组织样本用 10% 福尔马林固定, 石蜡包埋, 连续冠状切片, 厚度为 4-6 μ m。切片 70°C 加热 10 min, 之后依次浸泡于下述溶液中: 二甲苯 (5 min), 96% 乙醇 (3 min), 90% 乙醇 (3 min), 双蒸水 (3 min)。切片经 nesfatin-1 (1:2000, 美国 Phoenix 公司) 或 ghrelin (1:10000, 美国 sigma 公司) — 抗孵育, 4°C 过夜^[11], 磷酸盐缓冲液清洗后, 用羊抗兔生物素 Ig G 孵育 4°C 过夜^[11]。切片经卵白素 - 生物素 - 过氧化物酶复合物着色 10 min^[11], 梯度乙醇溶液中脱水, 苏木紫中性染色。光镜下观察。免疫活性细胞密度(D)用以下公式计算:

$$D = (Ng/Nt) \times 100\%$$

Ng: 一定区域免疫活性细胞的数量; Nt: 免疫活性细胞的总数量。

1.5 统计学分析

所有数据均用 ($\bar{x} \pm s$) 表示, 多组间均数比较采用单因素方差分析, 两组间均数比较采用 t 检验, Real-time PCR 结果分析采用 ΔCt 值表示, 即检测待测基因 cDNA 进入 PCR 指数增长期的起始点即循环阈值 (cycle threshold, Ct), 以同一样本中神经肽和看家基因 β -actin 之间的 Ct 差值 (ΔCt), 即 $\Delta Ct = Ct_{\text{ghrelin}} - Ct_{\beta\text{-actin}}$ 进行组间资料的 t 检验 (Prism 3.0 统计软件), $P<0.05$ 为差异有显著统计学意义。

2 结果

与对照组相比, 糖尿病大鼠体重显著降低 ($t=23.16$, $P<0.01$; 表 1), 糖尿病大鼠胃湿重与对照组大鼠比较无明显差异 ($t=0.68$, $P>0.05$)。血糖水平显著升高 ($t=22.55$, $P<0.01$), 血浆胰岛素和 IGF-1 水平显著降低 ($t=6.50$, $t=24.13$, $P<0.01$), 但 GH 水平显著升高 ($t=3.30$, $P<0.05$)。

与对照组大鼠相比, 糖尿病大鼠血浆总 ghrelin ($t=7.03$, $P<0.01$) 和活性 ghrelin ($t=3.13$, $P<0.05$) 水平均显著升高, 而 nesfatin-1 ($t=6.24$, $P<0.01$) 水平则显著降低 (表 2)。糖尿病大鼠血浆总 ghrelin 水平与 GH 水平 ($r=0.81$, $P<0.01$) 和 IGF-1 水平 ($r=-0.58$, $P<0.01$) 呈显著相关性 (图 1A,B)。与对照组大鼠相比, 糖尿病大鼠胃总 ghrelin ($t=16.86$, $P<0.01$) 和活性 ghrelin ($t=3.30$, $P<0.05$) 含量均显著降低, 而胃 nesfatin-1 含量则显著升高 ($t=7.93$, $P<0.01$, 表 2), 胃总 ghrelin 水平与血浆 IGF-1 水平呈明显相关性 ($r=0.65$, $P<0.01$, 图 1C)。

表 1 糖尿病大鼠体重、血糖、胃湿重及血浆胰岛素和 GH 水平($\bar{x} \pm s$)Table 1 The levels of body weight, gastric weight, serum glucose, serum insulin, serum IGF-1 and serum GH in STZ-induced diabetic rats ($\bar{x} \pm s$)

	Body weight (g)	Gastric weight(mg)	Glucose (mg/dL)	Insulin (ng/mL)	GH (ng/mL)	IGF-1 (ng/mL)
Control	353.4± 16.0	2145.6± 81.2	136.5± 4.1	0.27± 0.08	6.63± 0.24	1478.6± 137.6
Diabetic	207.5± 12.2**	2160.5± 85.0	451.2± 41.5**	0.08± 0.05**	35.23± 13.24*	449.5± 32.5**

注: *P<0.05, **P<0.01, 与对照组相比。

Notes: *P<0.05, **P<0.01 vs. control group.

表 2 糖尿病大鼠血浆和胃组织 ghrelin 和 nesfatin-1 的表达($\bar{x} \pm s$)Table 2 The level of plasma ghrelin, nesfatin-1 and gastric ghrelin, nesfatin-1 in STZ-induced diabetic rats ($\bar{x} \pm s$)

Group	Total Ghrelin		Active Ghrelin		Nesfatin-1	
	Plasma (fmol/mL)	Gastric (fmol/mg)	Plasma (fmol/mL)	Gastric (fmol/mg)	Plasma (fmol/mL)	Gastric (fmol/mg)
	(fmol/mL)	(fmol/mg)	(fmol/mL)	(fmol/mg)	(fmol/mL)	(fmol/mg)
Control (n=15)	615.2± 72.3	1758.8± 184.3	76.2± 25.6	337.5± 25.8	357.8± 53.1	1132.5± 96.5
Diabetic (n=15)	1123.4± 196.6**	1007.5± 195.2**	120.2± 38.5*	205.2± 36.5**	216.2± 48.3*	1385.5± 103.3*

注: *P<0.05, **P<0.01, 与对照组相比。

Notes: *P<0.05, **P<0.01 vs. control group.

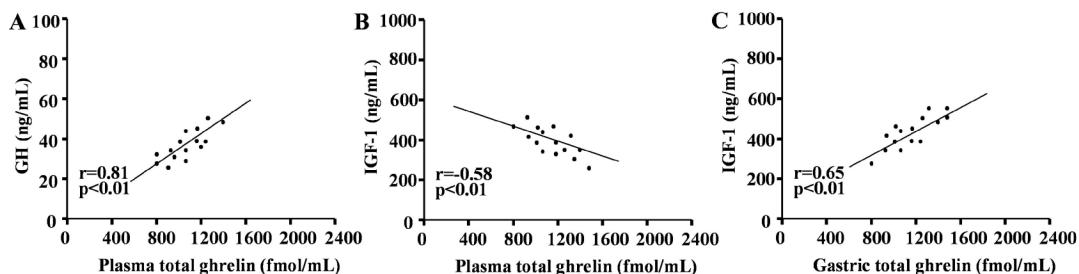


图 1 Ghrelin 与 GH, IGF-1 水平的相关性分析

Fig. 1 The correlation between ghrelin and GH, IGF-1

注: A: 血浆总 ghrelin 与 GH 水平的相关性分析; B: 血浆总 ghrelin 与 IGF-1 水平的相关性分析; C: 胃总 ghrelin 与 IGF-1 水平的相关性分析。

Note: A: The correlation between plasma total ghrelin and GH; B: The correlation between plasma total ghrelin and IGF-1; C: The correlation between gastric total ghrelin and IGF-1.

与对照组相比, 糖尿病大鼠胃 ghrelin mRNA 表达水平显著升高($t=16.8, P<0.01$; 图 2a), 但十二指肠和结肠 ghrelin mRNA 表达水平与对照组无显著差异($t=1.24, P>0.05$)。与对照组相比, 糖尿病大鼠胃底 ghrelin 免疫活性细胞数量显著减少

($t=3.98, P<0.01$; 图 2b), 胃底 ghrelin 免疫活性细胞与细胞总数比例减小, 提示糖尿病大鼠胃非活性细胞增加, 但与对照组相比差异不明显($t=1.64, P=0.11$; 图 2c)

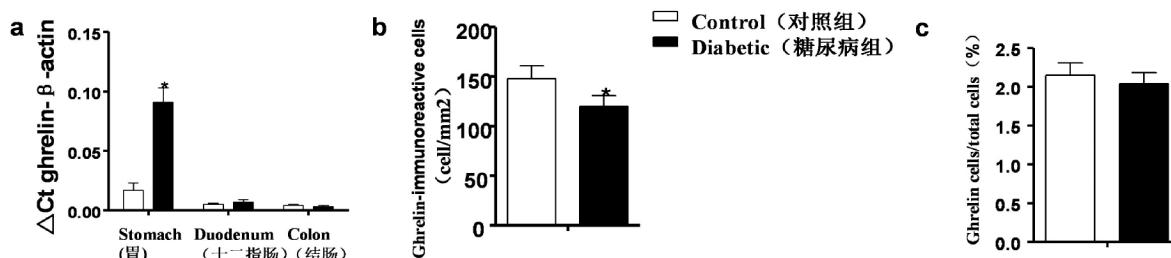


图 2 糖尿病大鼠 ghrelin mRNA 和 ghrelin 免疫反应细胞的表达

Fig. 2 The expression of preproghrelin mRNA and ghrelin-immunoreactive cells in STZ-induced diabetic rats

注: a: 胃、十二指肠和结肠中 ghrelin mRNA 的表达(ghrelin/β -actin); b: 糖尿病大鼠 ghrelin 免疫活性细胞数量;

c: 胃底 ghrelin 免疫活性细胞与细胞总数比例。*P<0.01, 与对照组相比

Note: a: the expression of ghrelin mRNA in stomach, duodenum and colon (ghrelin/β -actin); b: the number of ghrelin-immunoreactive cells of diabetic rats; c: the ratio of ghrelin-immunoreactive cells in total cells in fundus of stomach. *P<0.01 vs. control group.

实验中给予大鼠自由饮食,糖尿病大鼠血浆总 ghrelin 水平显著增加 ($t=7.53, P<0.01$; 表 3), nesfatin-1 水平显著降低 ($t=5.46, P<0.01$)。胰岛素处理后可使增加的 ghrelin 水平降低接近正常 ($t=1.76, P=0.11$), 而使降低的 nesfatin-1 水平升高 ($t=1.96, P=0.06$)。与对照组相比,糖尿病大鼠胃总 ghrelin 水平降低 ($t=5.14, P<0.01$), 而 nesfatin-1 水平显著升高 ($t=5.37, P<0.01$), 给予胰岛素可显著反转 ghrelin ($t=8.54, P<0.01$; 表 3) 和

nesfatin-1 水平 ($t=2.42, P<0.05$)。在正常大鼠、糖尿病大鼠和胰岛素处理糖尿病大鼠之间,胃内活性 ghrelin 与总 ghrelin 的比率无显著差异 ($t=1.12, P>0.05$)。与正常大鼠比较,糖尿病大鼠胃底 ghrelin 免疫活性细胞数显著减少 ($t=3.56, P<0.01$), nesfatin-1 免疫活性细胞数量明显增加 ($t=3.56, P<0.01$), 胰岛素处理后可显著改善这一状况 ($t=3.21, t=2.59, P<0.05$; 表 3)。

表 3 胰岛素对糖尿病大鼠 ghrelin 和 nesfatin-1 表达的影响 ($\bar{x}\pm s$)Table 3 The effects of insulin on expression of ghrelin and nesfatin-1 in STZ-induced diabetic rats ($\bar{x}\pm s$)

Group	Plasma total ghrelin (fmol/mL)	Gastric total ghrelin (fmol/mg)	Ghrelin Cells (Cell/mm ²)	Plasma nesfatin-1 (fmol/mL)	Gastric nesfatin-1 (fmol/mg)	Nesfatin-1 Cells (Cell/mm ²)
Control (n=15)	492.1± 43.2	2238.1± 197.5	96.2± 12.5	275.5± 41.3	1252.3± 114.0	48.5± 9.6
Diabetic (n=15)	714.4± 58.4**	1652.1± 158.2**	61.7± 10.3**	182.6± 35.8**	1537.4± 123.1**	64.4± 11.3**
Diabetic+INS (n=15)	655.3± 52.5	2690.5± 204.5#	108.2± 22.5#	214.4± 38.6	1406.2± 119.2#	50.1± 6.5

注: **P<0.01, 与对照组相比; #P<0.05, ##P<0.01, 与糖尿病组相比。

Notes: **P<0.01 vs. control group; #P<0.05, ##P<0.01 vs. diabetic group.

3 讨论

STZ 处理大鼠体重和血浆胰岛素水平显著降低,血糖水平显著升高。这种观点与之前研究结果一致 [13], 提示本实验中 STZ 诱导糖尿病大鼠模型成功。大鼠给予外源性 ghrelin 可增加血浆 GH 水平 [16], STZ 诱导的糖尿病大鼠血浆 GH 水平与 ghrelin 水平有显著相关性,与在恶病质和慢性心脏病中研究相符[6]。STZ 诱导糖尿病大鼠研究结果显示,血浆 ghrelin 水平升高,胃 ghrelin mRNA 表达水平降低,胃底 ghrelin 水平与 ghrelin 免疫活性细胞数量显著降低,而血浆 nesfatin-1 水平显著降低,胃 nesfatin-1 水平显著升高。有文献报道,十二指肠有 ghrelin、ghrelin mRNA 表达,ghrelin 免疫活性细胞数量也较多 [11,17,18], 本研究显示结肠和十二指肠 ghrelin mRNA 表达水平无显著差异,提示,ghrelin 主要在胃中产生。且有报道 nesfatin-1 在胃组织中的表达是脑的 10 倍。提示糖尿病大鼠胃内 ghrelin 水平降低,nesfatin-1 水平升高可能与血浆 ghrelin 水平的升高, nesfatin-1 水平的降低有重要关联。

Ghrelin 和 nesfatin-1 均可由胃 X/A 样细胞分泌 [19]。本研究发现,胰岛素处理大鼠可逆转胃底 ghrelin 和 nesfatin-1 免疫活性细胞数量的改变,提示胃 A 样细胞中 ghrelin 或 nesfatin-1 标记细胞数量改变,而不是 A 样细胞分泌 ghrelin 或 nesfatin-1 变化。糖尿病大鼠血浆 ghrelin 水平升高,胃 ghrelin 水平及 ghrelin 免疫活性细胞数量降低可能归因于胃 ghrelin 释放入血增多相关。糖尿病大鼠体重显著降低,胰岛素处理后可逆转糖尿病大鼠 ghrelin 或 nesfatin-1 动力学改变,这些改变可能导致负能量平衡,如体重降低,而不是直接逆转 STZ 对胃粘膜毒性作用。但对刺激肽类合成或分泌的信号通路机制还不清楚。

Ghrelin 是一种具有多重生理功能的脑肠肽,研究显示, ghrelin 与 IGF-1 相似具有双重调控代谢和生长的作用 [20]。本研究结果发现,血浆 IGF-1 水平与血浆和胃总 ghrelin 水平有显著相关性,血浆 IGF-1 水平的降低可能参与这一调控机制。另

外,负能量平衡状态可能引起胃 ghrelin mRNA 表达水平补偿性上调,使 ghrelin 合成分泌增加。

总之,STZ 诱导糖尿病大鼠胃 ghrelin 分泌入血增加,提示,在 STZ 处理大鼠,负能量平衡可能导致 ghrelin mRNA 水平上调,从而刺激胃 ghrelin 前体细胞合成分泌 ghrelin,而胃 nesfatin-1 水平升高可能与糖尿病胃轻瘫的发生发展有相关性。

参考文献(References)

- [1] Kojima M, Hosoda H, Date Y, et al. Ghrelin is a growth-hormone-releasing acylated peptide from stomach [J]. Nature, 1999, 402(7): 656-660
- [2] Bilski J, Mańko G, Brzozowski T, et al. Effects of exercise of different intensity on gut peptides, energy intake and appetite in young males[J]. Ann Agric Environ Med, 2013, 20(4): 787-793
- [3] Masuda Y, Tanaka T, Inomata N, et al. Ghrelin stimulates gastric acid secretion and motility in rats [J]. Biochem Biophys Res Commun, 2000, 276(32): 905-908
- [4] Di Fonzo A, Ghinassi B, Izzicupo P, et al. Novel evidence of ghrelin and growth hormone secretagogue receptor expression by human ocular tissues[J]. Regul Pept, 2014, 190-191C:18-24
- [5] Date Y, Murakami N, Toshinai K, et al. The role of the gastric afferent vagal nerve in ghrelin-induced feeding and growth hormone secretion in rats[J]. Gastroenterology, 2002, 123(4): 1120-1128
- [6] Invernizzi M, Carda S, Cisari C, et al. Possible synergism of physical exercise and ghrelin-agonists in patients with cachexia associated with chronic heart failure [J]. Aging Clin Exp Res, 2013, 18(23): 75-86
- [7] Asakawa A, Inui A, Kaga T, et al. Ghrelin is an appetite-stimulatory signal from stomach with structural resemblance to motilin [J]. Gastroenterology, 2001, 120(2): 337-345
- [8] Mé quinon M, Langlet F, Zgheib S, et al. Ghrelin: central and peripheral implications in anorexia nervosa [J]. Front Endocrinol, 2013, 26(7): 4-15
- [9] Wei R, Liu T, Zhou C, et al. Identification, tissue distribution and

- regulation of preproghrelin in the brain and gut of Schizothorax prenanti [J]. Gastroenterology, 2013, 10(186): 18-25
- [10] Kim HH, Jeon TY, Park do Y, et al. Differential expression of ghrelin mRNA according to anatomical portions of human stomach [J]. Hepatogastroenterology, 2012, 59(119): 2217-2221
- [11] Oh-I S, Shimizu H, Satoh T, et al. Identification of NUCB2/nesfatin-1 as a satiety molecule in the hypothalamus [J]. Nature, 2006, 443(7112): 709-712
- [12] Szkudelski T, Zywert A, Szkudelska K, et al. Metabolic disturbances and defects in insulin secretion in rats with streptozotocin-nicotinamide-induced diabetes [J]. Physiol Res, 2013, 62(6): 663-670
- [13] Wang L, Duan G, Lu Y, et al. The effect of simvastatin on glucose homeostasis in streptozotocin induced type 2 diabetic rats [J]. J Diabetes Res, 2013, 20(13): 274-286
- [14] Bang AS, Soule SG, Yandle TG, et al. Characterisation of proghrelin peptides in mammalian tissue and plasma [J]. J Endocrinol, 2007, 192(2): 313-323
- [15] Prudom C, LIU J, Patrice J, et al. Comparison of competitive radioimmunoassays and two-site sandwich assays for the measurement and interpretation of plasma ghrelin levels [J]. J Clin Endocrinol Metab, 2010, 95(5): 2351-2358
- [16] Inoue H, Sakamoto Y, Kangawa N, et al. Analysis of expression and structure of the rat GH-secretagogue/ghrelin receptor (Ghsr) gene: roles of epigenetic modifications in transcriptional regulation [J]. Mol Cell Endocrinol, 2011, 345(12): 1-15
- [17] Kasacka I, Arciszewski M, Lebkowski W, et al. Extraordinary level of hormone and number of ghrelin cells in the stomach and duodenum of an obese woman [J]. Acta Histochem, 2014, 116(1): 230-234
- [18] Teive MB, Russi RF, Vieira DS, et al. Quantitative immunohistochemical analysis of duodenal ghrelin cells after sleeve gastrectomy in Wistar rats [J]. Acta Cir Bras, 2012, 27(9): 595-599
- [19] Andreas S, Yvette T, Yin and Yang-the Gastric X/A-like cell as possible dual regulator of food intake [J]. J Neurogastroenterol Motil, 2012, 18(2): 2087-2093
- [20] Arellanes-Licea Edel C, Báez-Ruiz A, Carranza ME, et al. Daily patterns and adaptation of the ghrelin, growth hormone and insulin-like growth factor-1 system under daytime food synchronisation in rats [J]. J Neuroendocrinol, 2014, 26(5): 282-295

(上接第 1811 页)

- [5] 冯丹妮, 陶虹, 德向研, 等. 癫痫致大鼠岛叶皮层及海马 GABA 能神经元亚型变化的形态学观察 [J]. 宁夏医科大学学报, 2013, 35(8): 841-844, 849, 封 3
- Feng Dan-ni, Tao Hong, De Xiang-yan, et al. Morphological Changes of GABA Neuron Subtypes in Insular Cortex and Hippocampal induced by Epilepsy in Rats [J]. Journal of Ningxia Medical University, 2013, 35(8): 841-844, 849, e3
- [6] Horton JR, Sawada K, Nishibori M. Structural basis for inhibition of histamine N-methyltransferase by diverse drug [J]. Journal of Molecular Biology, 2005, (2): 334-344
- [7] Šapina L, Vuletić V, Lojen G, et al. Head trauma and posttraumatic epilepsy in Slavonski Brod, East Croatia, 1988-2008 [J]. Coll Antropol, 2014, 38(3): 1077-1079
- [8] Haas H, Panula P. The role of histamine and the tuberomamillary nucleus in the nervous system [J]. Nature Reviews Neuroscience, 2003, (2): 121-130
- [9] Suzuki R, Dickenson AH. Differential pharmacological modulation of the spontaneous stimulus-independent activity in the rat spinal cord following peripheral nerve injury [J]. Experimental Neurology, 2006, (1): 72-80
- [10] Mobarakeh JI, Takahashi K, Sakurada S. Enhanced antinociceptive effects of morphine in histamine H₂ receptor gene knockout mice [J]. Neuropharmacology, 2006, (3): 612-622
- [11] 黄亮, 足立尚登, 長格巧. 神经病理性疼痛大鼠纹状体组胺水平的变化及脑室内注射组胺的效果 [J]. 中华麻醉学杂志, 2006, (6): 542-546
- Huang Liang, Zu Li-shang deng, Chang Ge-qiao. Changes in histamine level in striatum in rats with neuropathic pain and the effects of intracerebral ventricular histamine on neuropathic pain [J]. Chinese Journal of Anesthesiology, 2006, (6): 542-546
- [12] Rajput SK, Singh JN, Ingole S, et al. Neuropharmacological profile of L-pGlu-(1-benzyl)-L-His-L-ProNH₂, a newer thyrotropin-releasing hormone analog: effects on seizure models, sodium current, cerebral blood flow and behavioral parameters [J]. Epilepsy Res, 2009, 87(2-3): 223-233
- [13] Rajput SK, Krishnamoorthy S, Pawar C, et al. Antiepileptic potential and behavioral profile of L-pGlu-(2-propyl)-L-His-L-ProNH₂, a newer thyrotropin-releasing hormone analog [J]. Epilepsy Behav, 2009, 14(1): 48-53
- [14] 成祥林, 赵成三, 汪华. 丙戊酸钠对戊四氮致痫大鼠海马 Bax 和 Bcl-2 表达的影响 [J]. 临床神经病学杂志, 2004, (04): 351-353
- Cheng Xiang-lin, Zhao Cheng-san, Wang Hua. Influence of the expression of Bax and Bcl-2 in hippocampus of sodium valproate on rat epileptic induced by E four nitrogen [J]. Journal of Clinical Neurology, 2004, (04): 351-353
- [15] Qu H, Eloqayli H, Sonnewald U. Pentylenetetrazole affects metabolism of astrocytes in culture [J]. Journal of Neuroscience Research, 2005, (1/2): 48-54
- [16] Shin S, Sung BJ, Cho YS. An anti-apoptotic protein human Survivin is a direct inhibitor of caspase-3 and caspase-7 [J]. Biochemistry, 2005, (04): 1117-1123
- [17] Rodríguez de la Vega RC, Schwartz Z EF, Possani LD. Mining on scorpion venom biodiversity [J]. Toxicology, 2010, (07): 1155-1161
- [18] Chippaux JP. Emerging options for the management of scorpion stings [J]. Drug Des Devel Ther, 2012: 165-173
- [19] Zhu J, Wang J, Cheng MS. Dinucleotides docking to scorpion polypeptide toxins: a molecular modeling method for protein functional site recognition [J]. Biochemical and Biophysical Research Communications, 2009, (02): 157-161
- [20] Jung S, Dingley AJ, Augustin R. Hydramacin-1, structure and antibacterial activity of a protein from the basal metazoan Hydra [J]. Journal of Biological Chemistry, 2009, (03): 1896-1905